

# ***Cell-free reaction platforms for multi-enzyme biocatalysis Challenges & Opportunities***

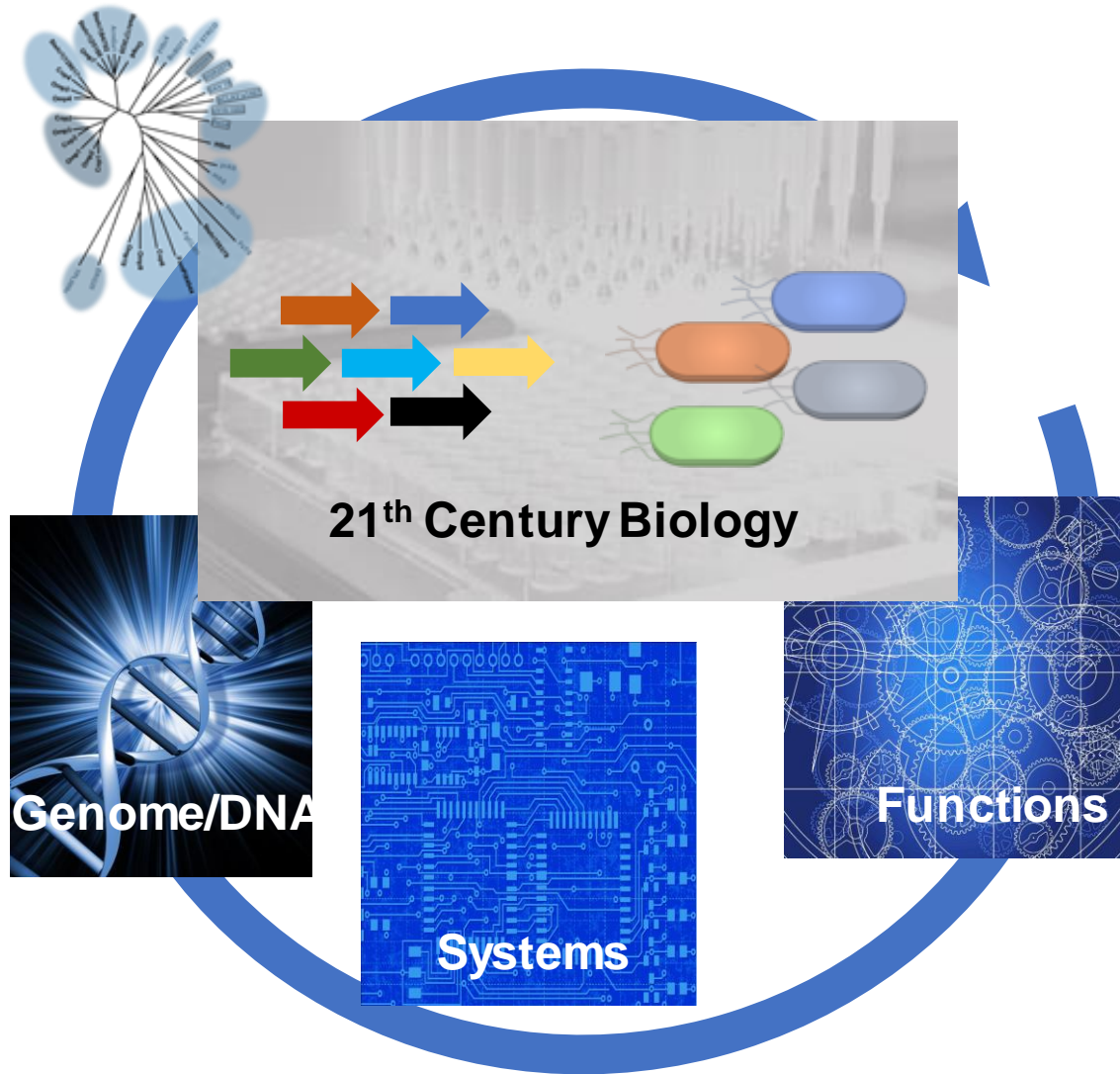
***Claudia Schmidt-Dannert***

*Dept. Biochemistry, Molecular Biology & Biophysics  
BioTechnology Institute*

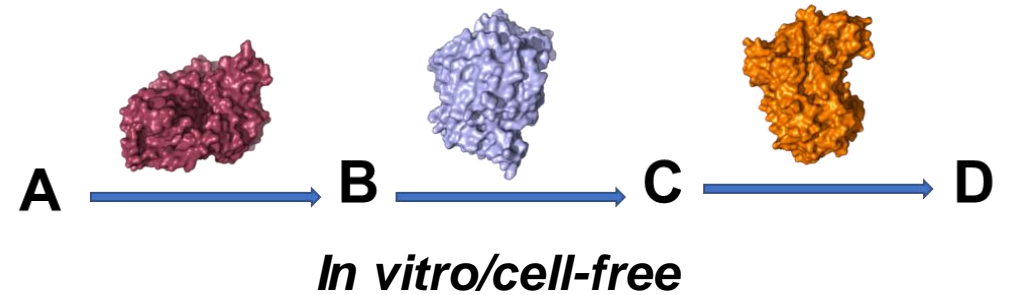
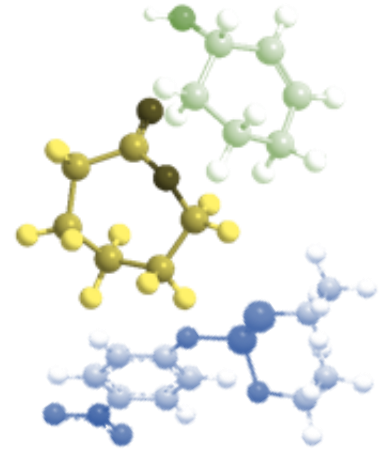


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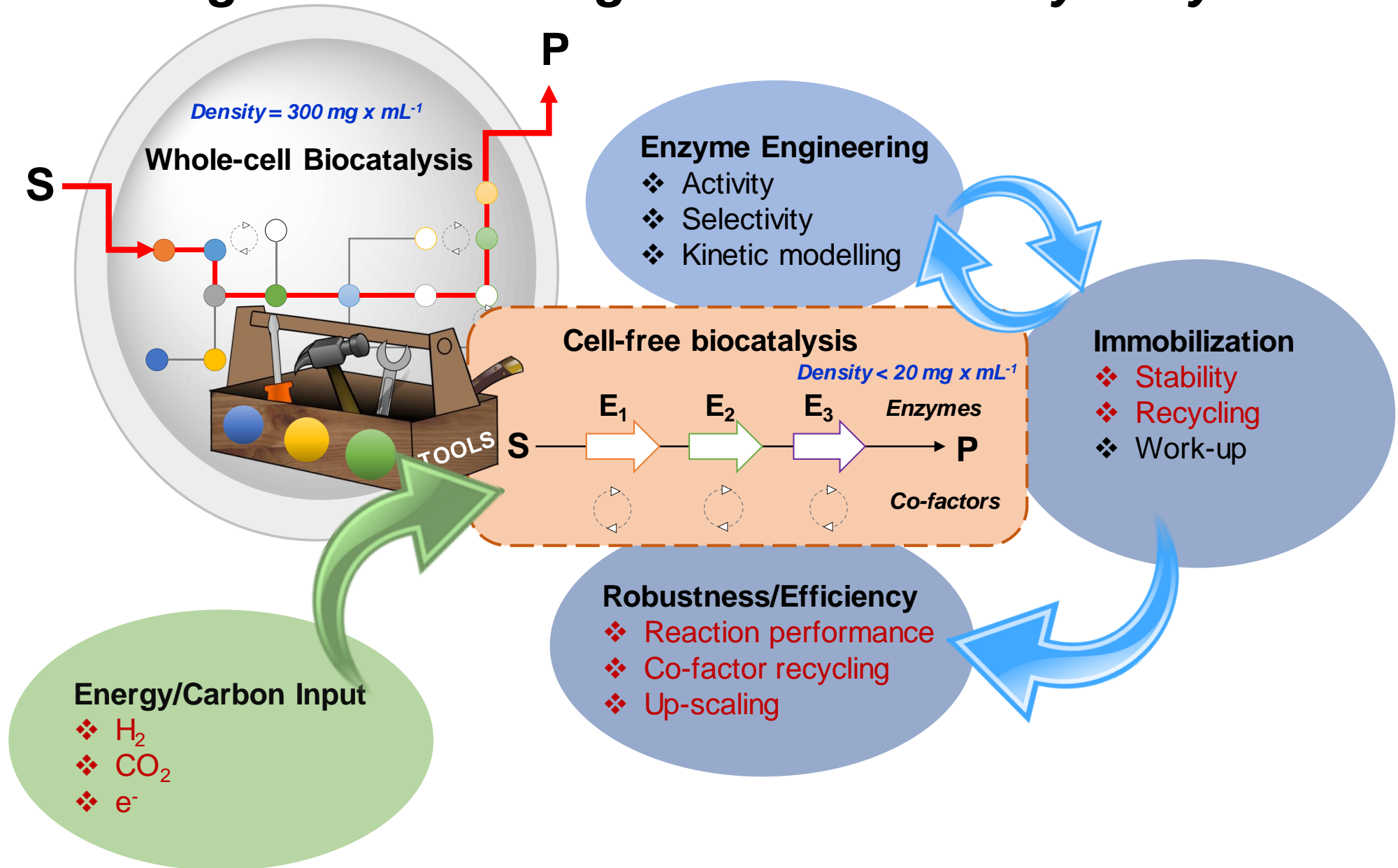
# Opportunities for the design of new biocatalytic systems



**Biomanufacturing**



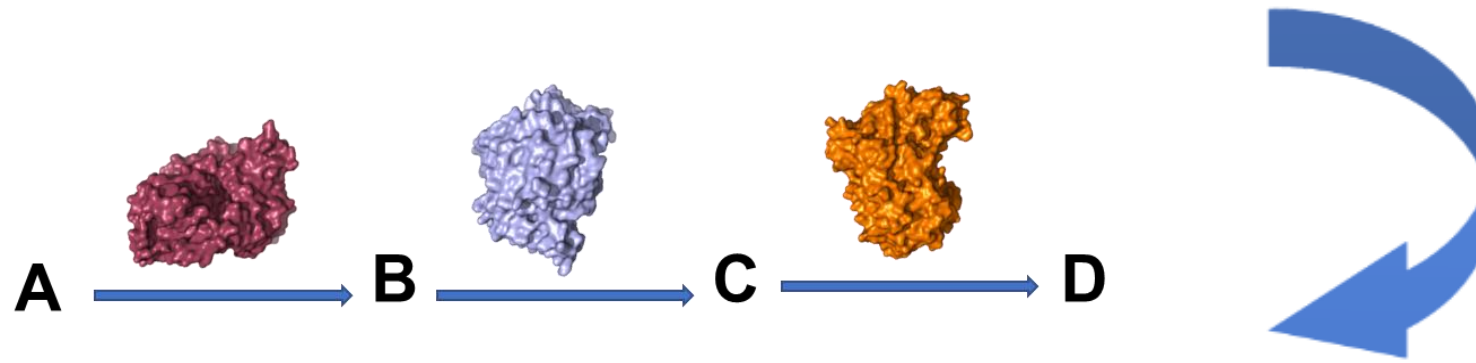
# Challenges for the design of new biocatalytic systems



# ***Fourth wave of biocatalysis\****

New enzyme classes for industrial biocatalysis

Rapid design of tailored enzyme reactions  
(Sequence/structure databases, HT-design)



Well-developed  
Amine synthesis  
Transaminases  
Ketoreductase/Alcohol DH  
Nitrilases

Emerging  
Imine reductases  
C-H Oxidations  
Aldolases

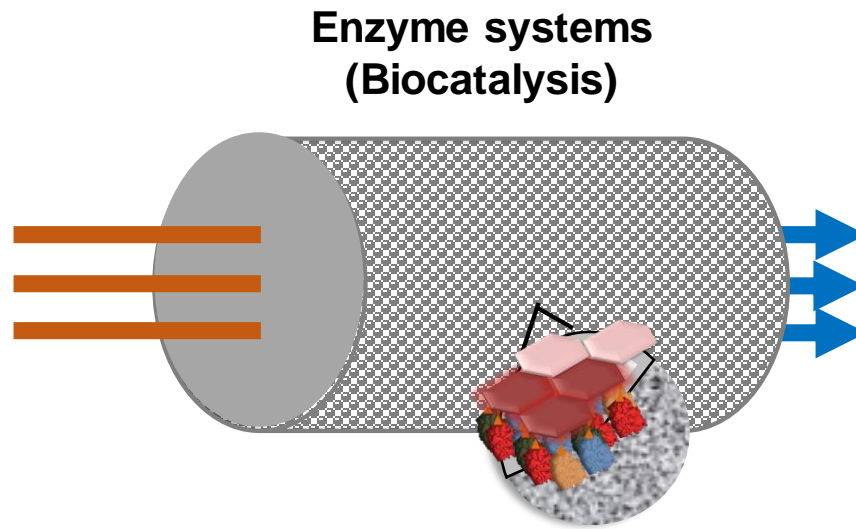
Large repertoire of reactions for the design of long enzyme cascades

New approaches for reaction and process design

Pharmaceuticals & Bulk Chemicals

# ***Merging synthetic biology and materials sciences***

## **Design of robust self-organized systems for biocatalysis**



**Biomanufacturing**

**Genetically programmable**

**De novo production**

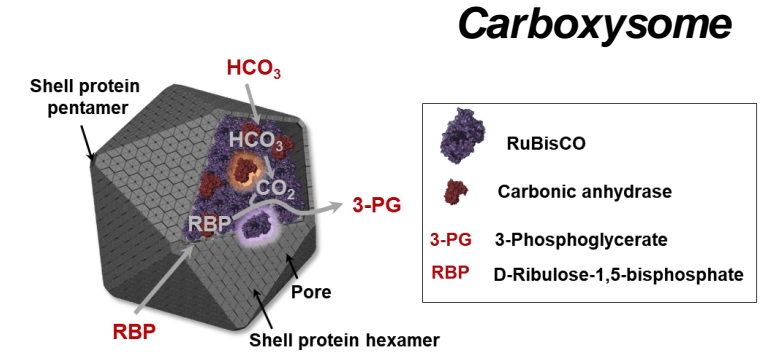
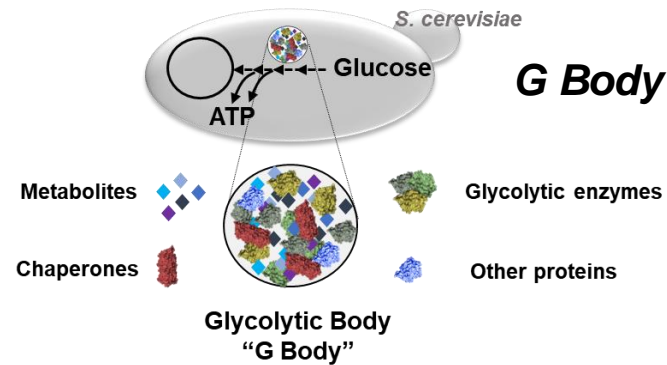
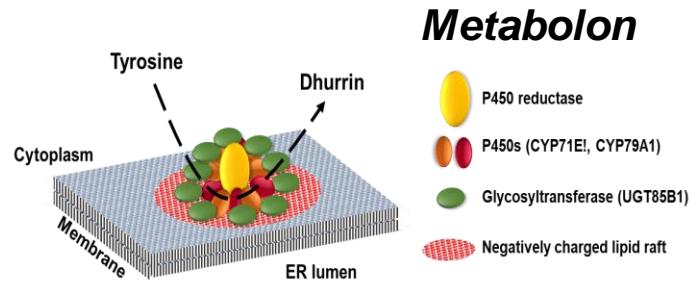
**Self-organization**

**Biomineralization**



# Self-organization in nature

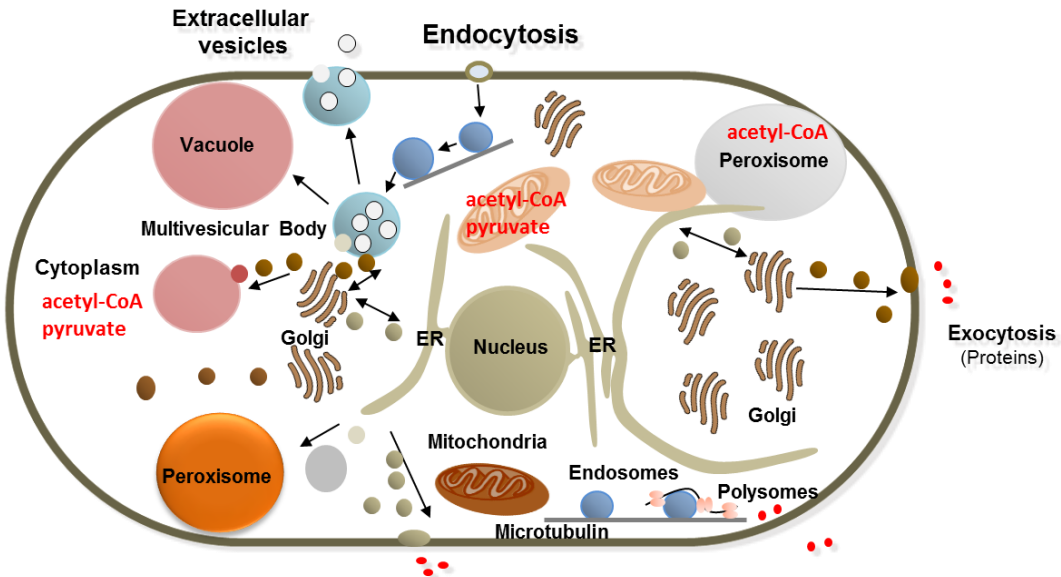
## Assembly of enzymes inside cells



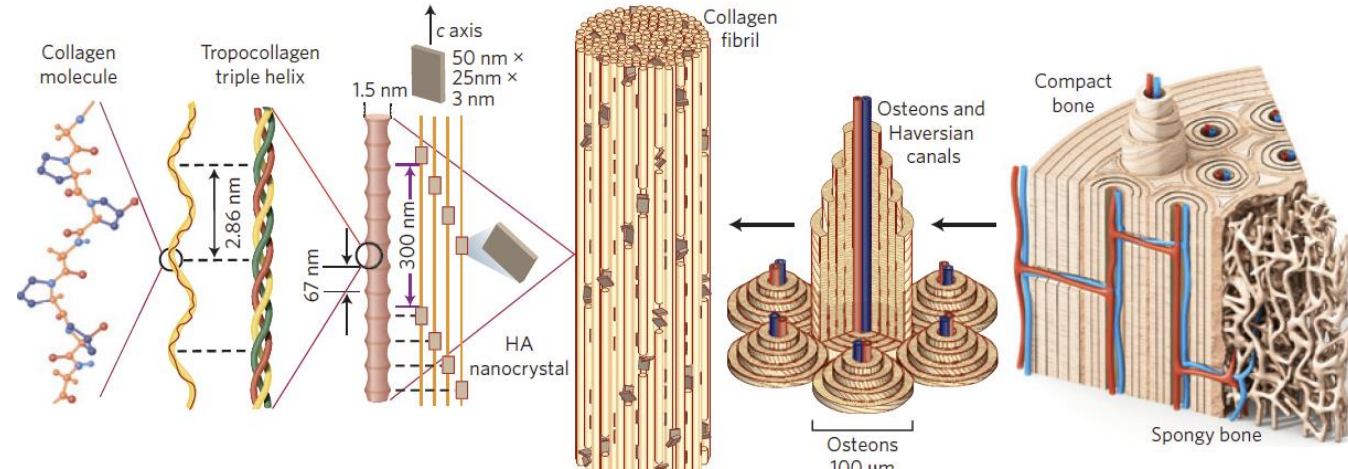
Bassard group: Science. 2016 354:890-893, Methods Enzymol. 2019 617:1-27

From: Schmidt-Dannert, Lopez-Gallego Curr. Opin. Chem. Biol., In Press

## Compartmentalization inside cells



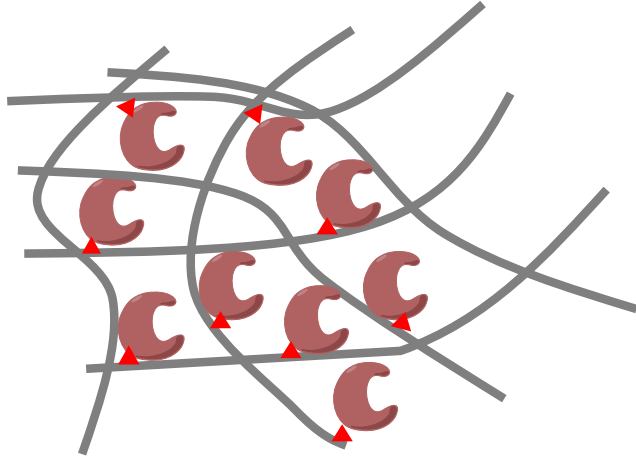
## Protein-templated higher ordered structures - Bone



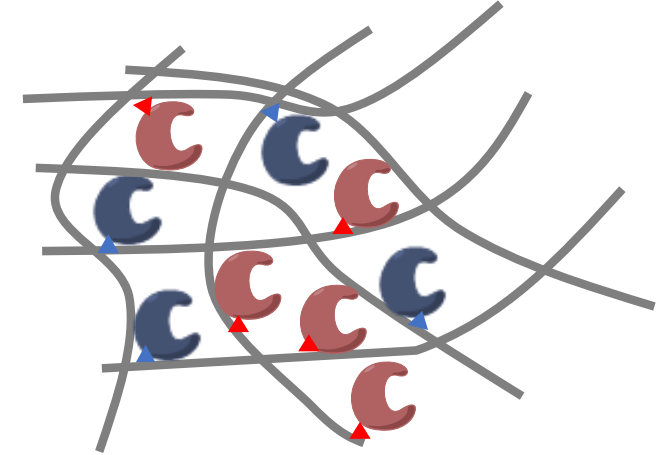
From: Nature Mat 2015 14:23-36

# Enzyme immobilization

## Enzyme stability & activity



## One-pot multi-enzyme catalysis



Challenges

- Enzyme compatible immobilization chemistries
- Operation of complex cascade reactions

## Manufacturing of enzyme system

Production



Isolation



Immobilization



Reuse

Scale-able

Cost-efficient

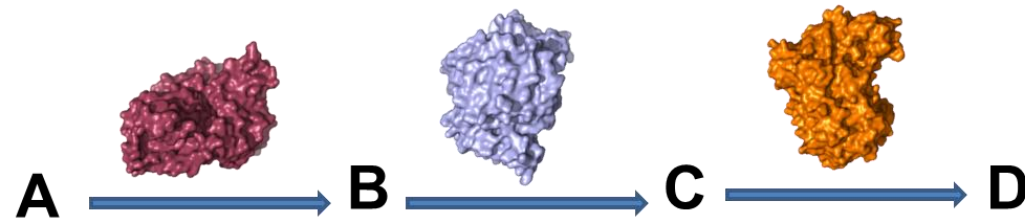
Cheap, mild, versatile

Robust & Efficient  
(High TOF & TON)

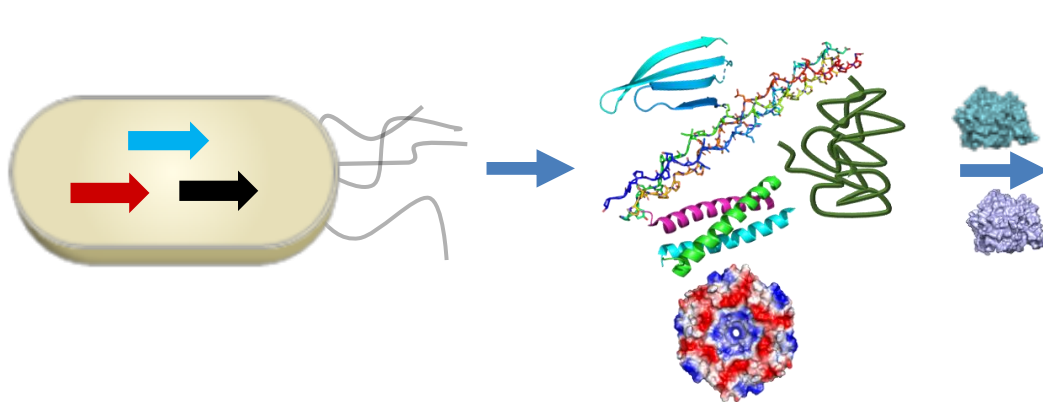
# ***Protein-based materials***

Development of structurally ordered biocomposite material with configurable (genetically programmable) material properties and embedded **biological capabilities**.

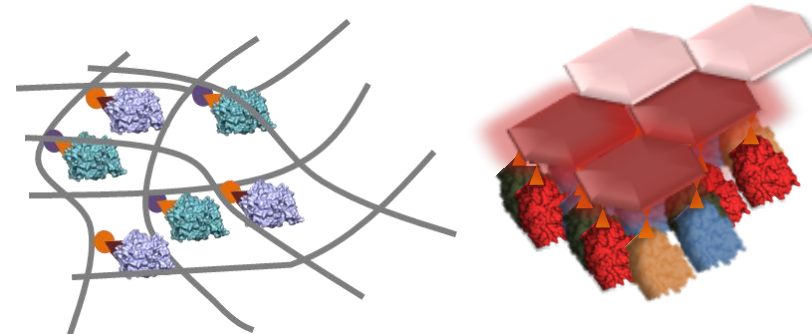
## ***Biocatalytic cascade***



## ***Configuration - Fabrication***

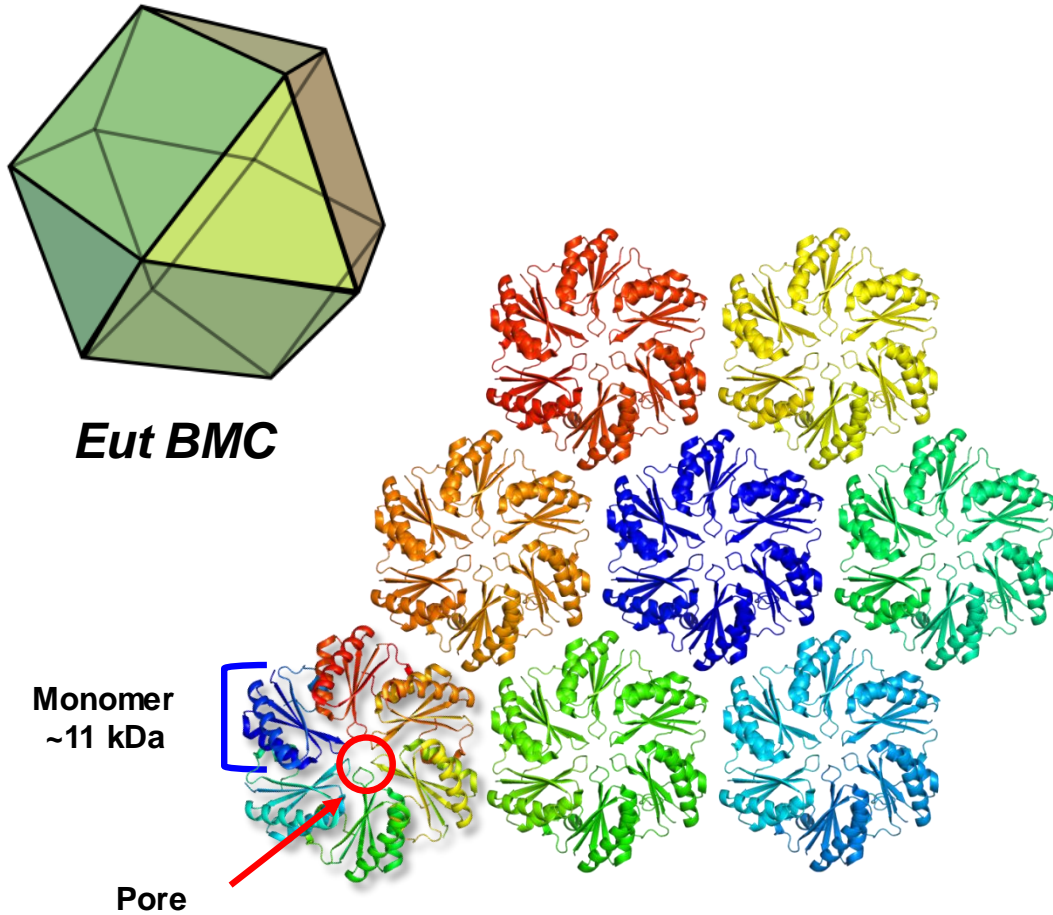


## ***Scaffolding - Immobilization - Operation***

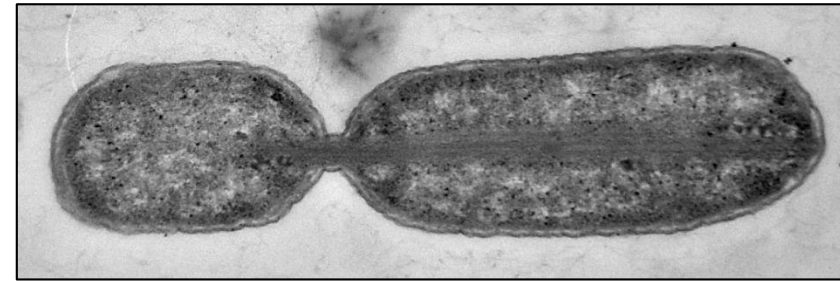




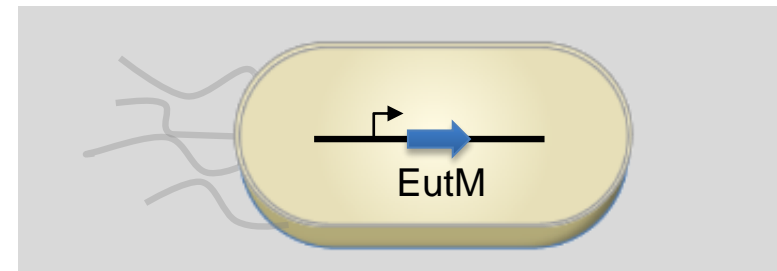
# Bacterial MicroCompartment shell protein as scaffold building block



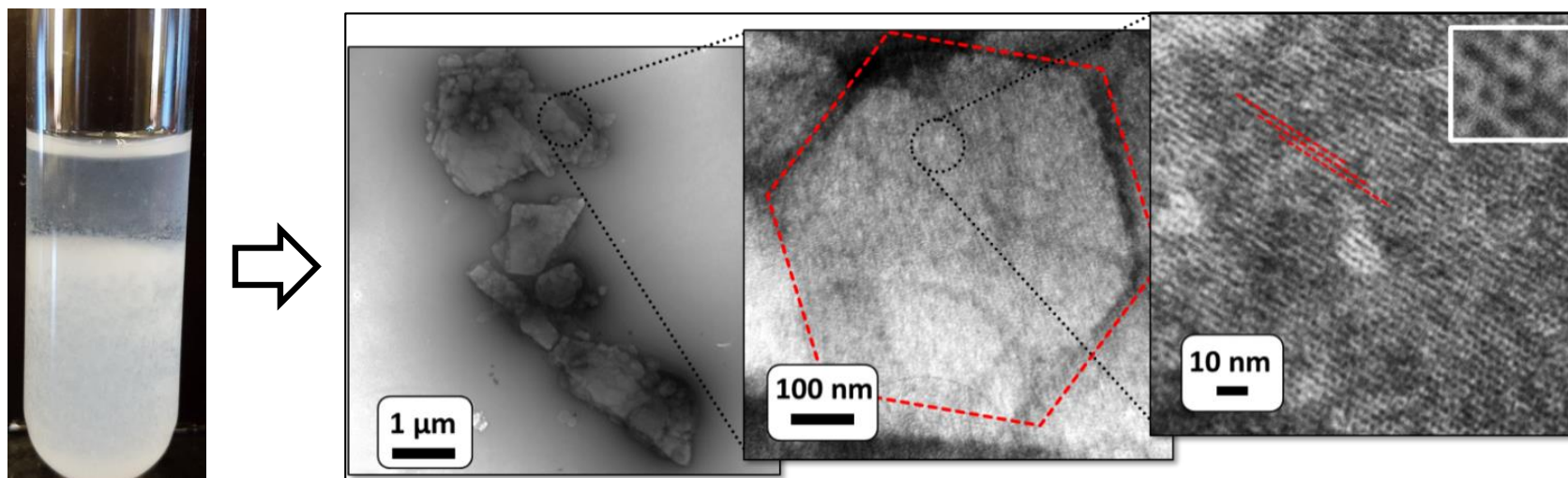
**EutM self-assembles as a hexameric crystal lattice**



**EutM expression in *E. coli* – protein scaffolds self-assemble *in vivo***

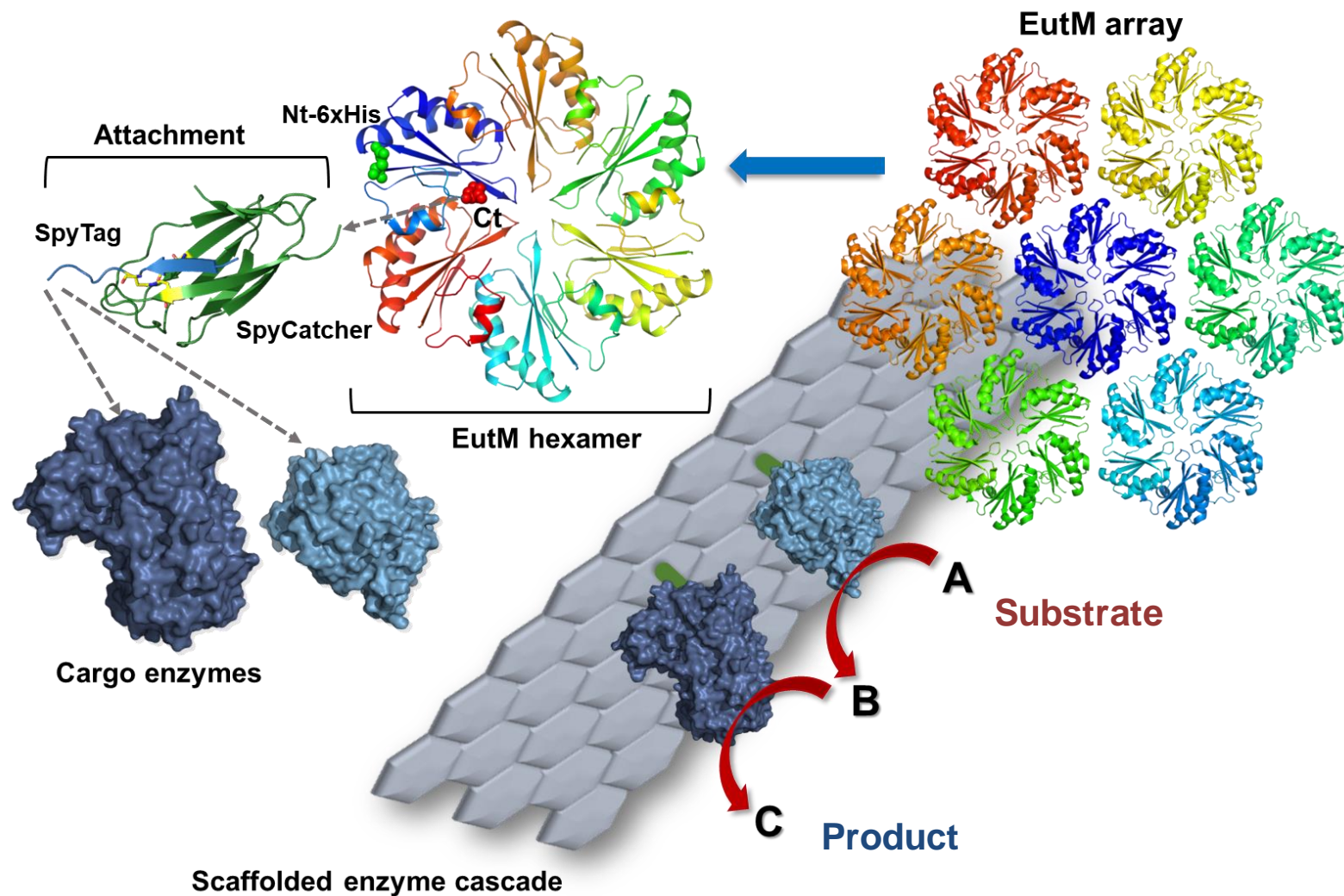


# ***Self-assembling into large scaffolds***



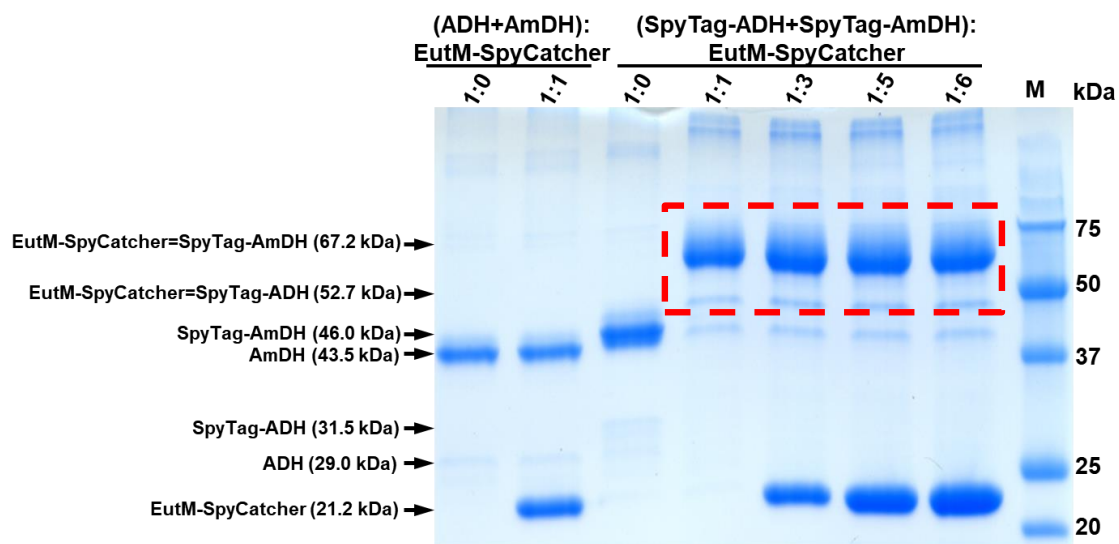
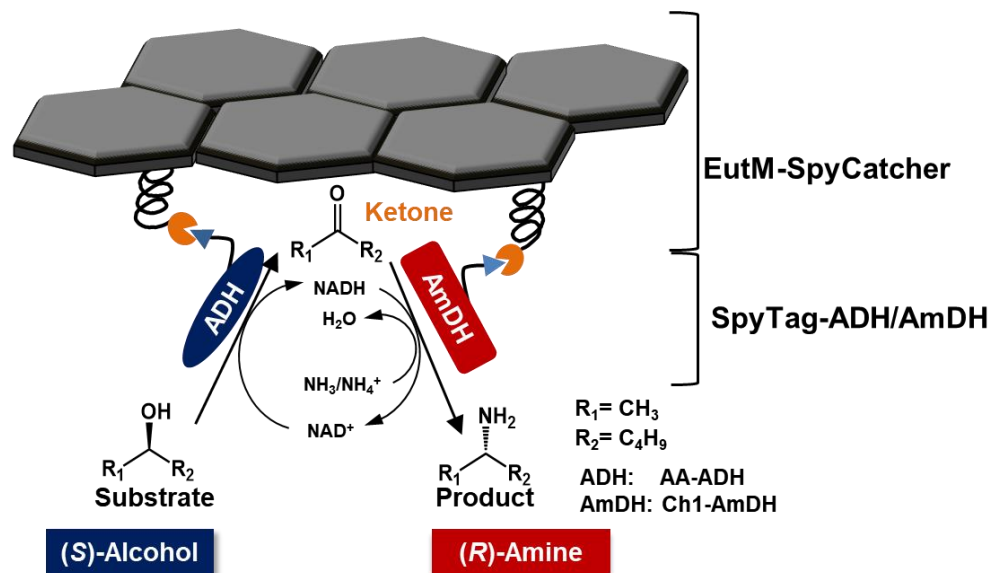
**EutM self-assembles *in vitro* as hexameric tiles and arrays**

# *Design of scaffolds for biocatalysis*



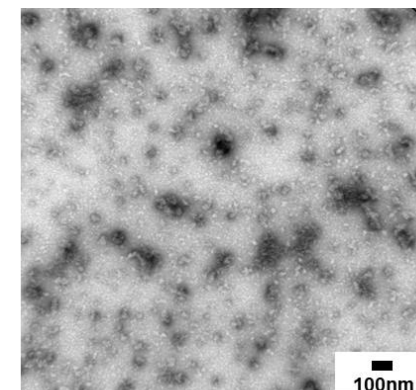


# Co-immobilization of enzyme cascade

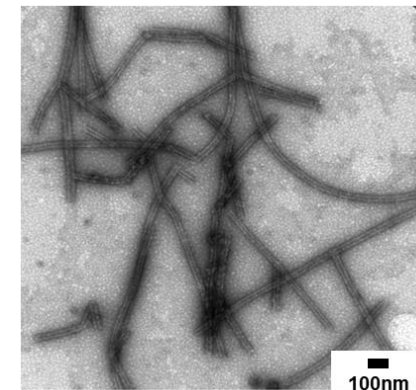


2 M ammonium chloride buffer pH 8.7  
(1:5 molar ratio enzymes:scaffold)  
6  $\mu\text{M}$  SpyTag-ADH  
150  $\mu\text{M}$  SpyTag-AmDH  
780  $\mu\text{M}$  EutM-SpyCatcher

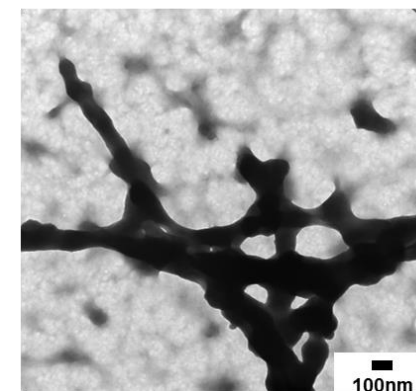
ACS Catalysis (2018) 8:5611-20



SpyTag-ADH + SpyTag-AmDH

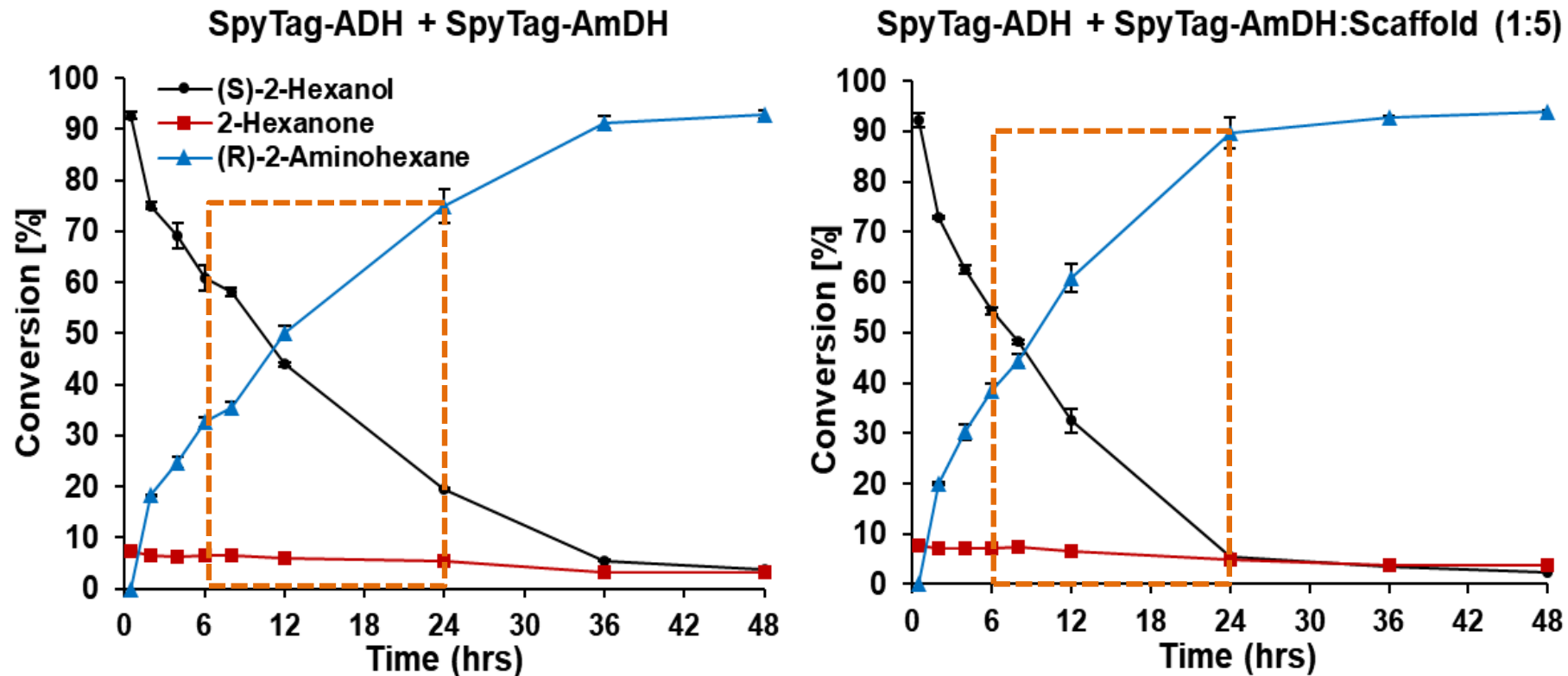


EutM-SpyCatcher



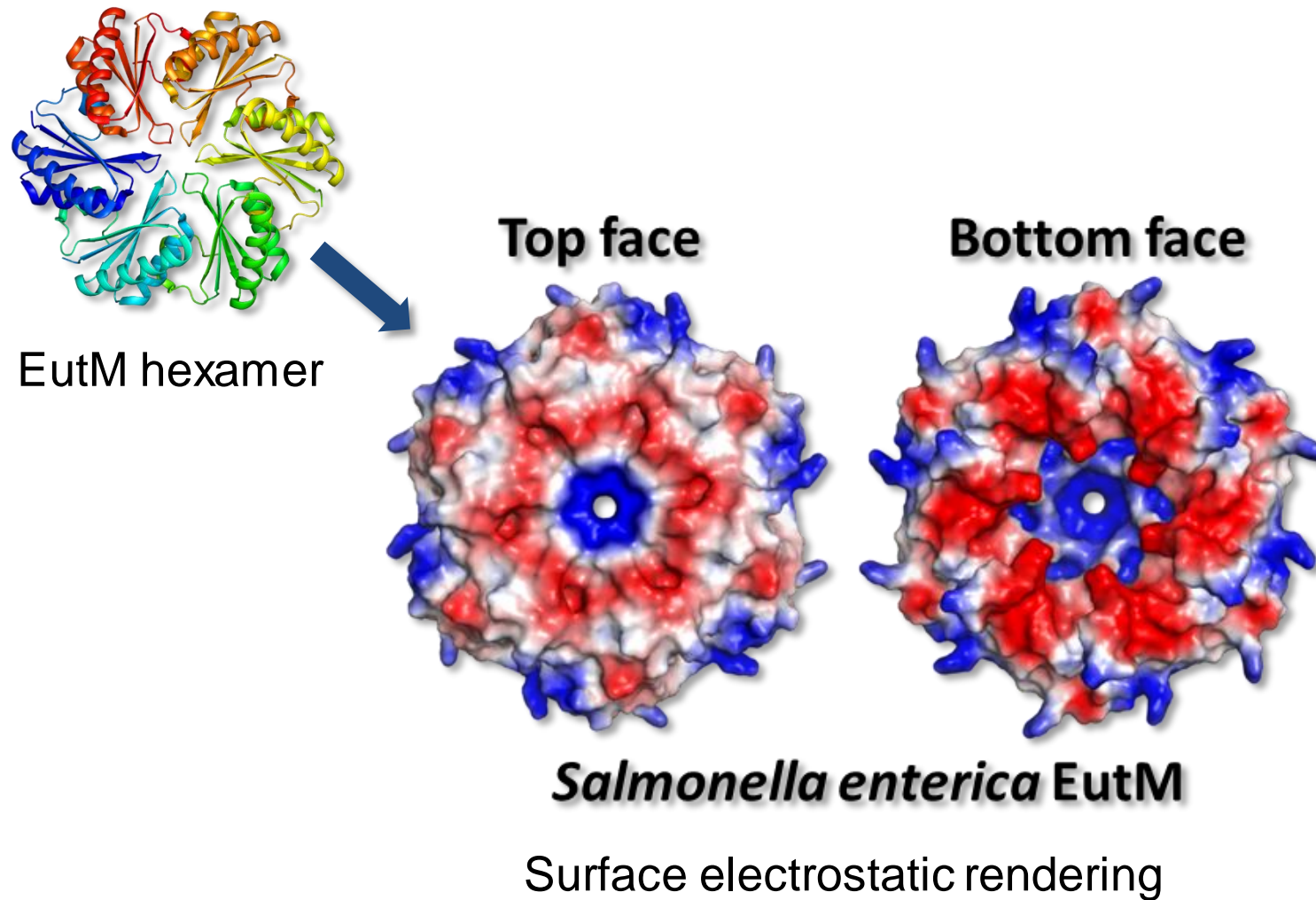
EutM-SpyCatcher +  
SpyTag-ADH + SpyTag-AmDH

# Optimization and testing of enzyme cascade



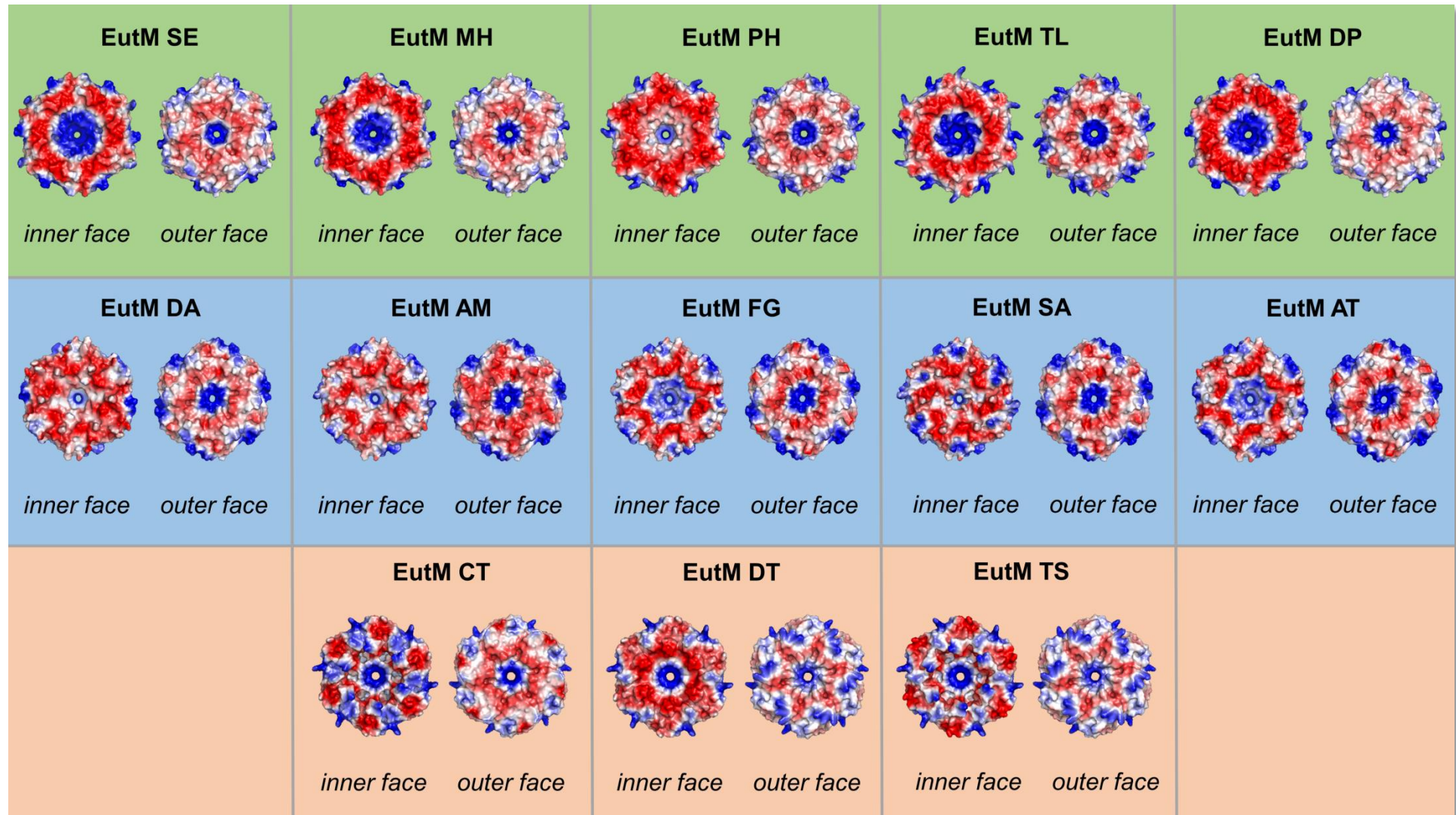
Scaffolds shorten reaction time to reach 90% conversion of 20 mM (S)-2-hexanol to (R)-2-aminohexane in 24 hrs with >99% ee.

# ***Scaffolds define electrostatics & architecture of materials***





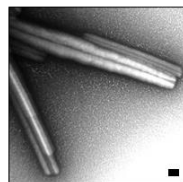
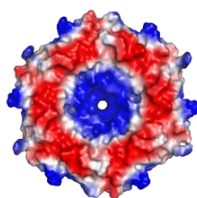
# *Expanding scaffold diversity*



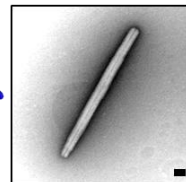
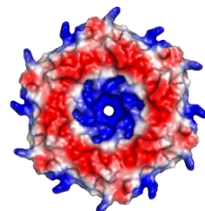
# *EutM-SpyCatcher building blocks*

## 8 EutM homologs selected

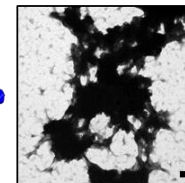
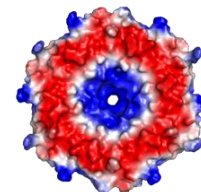
### Clade I



EutM SE

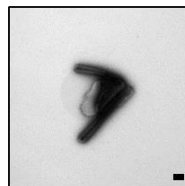
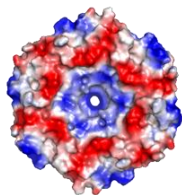


EutM TL

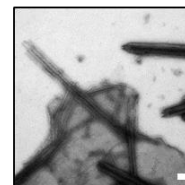
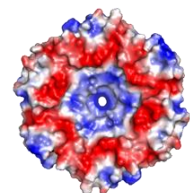


EutM DP

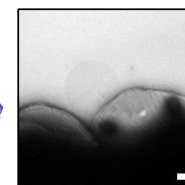
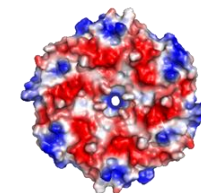
### Clade II



EutM AM

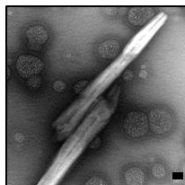
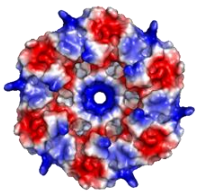


EutM FG

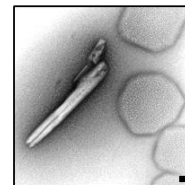
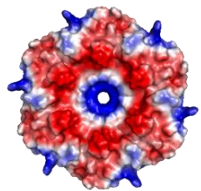


EutM SA

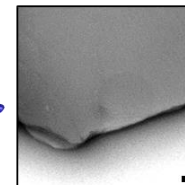
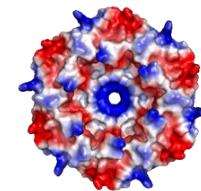
### Clade III



EutM CT

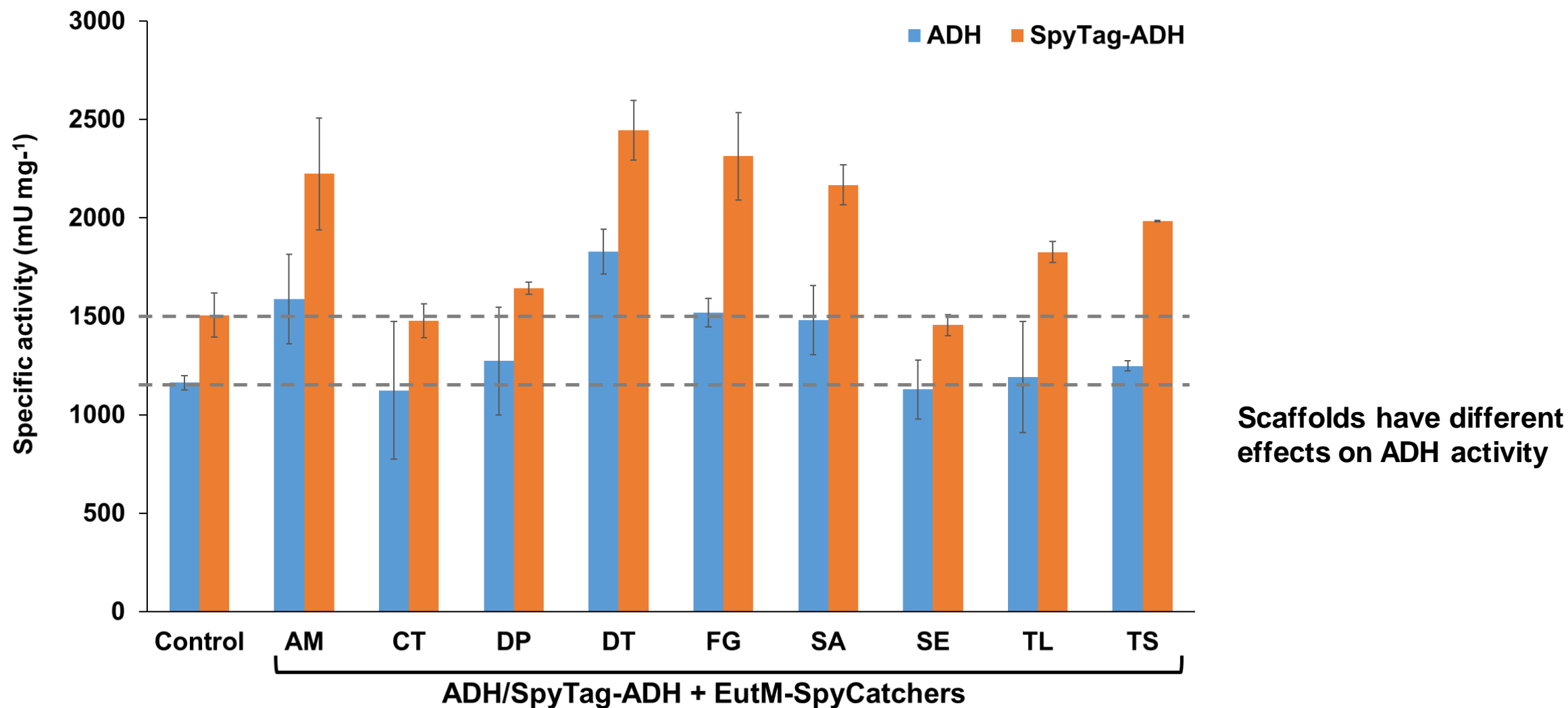


EutM DT



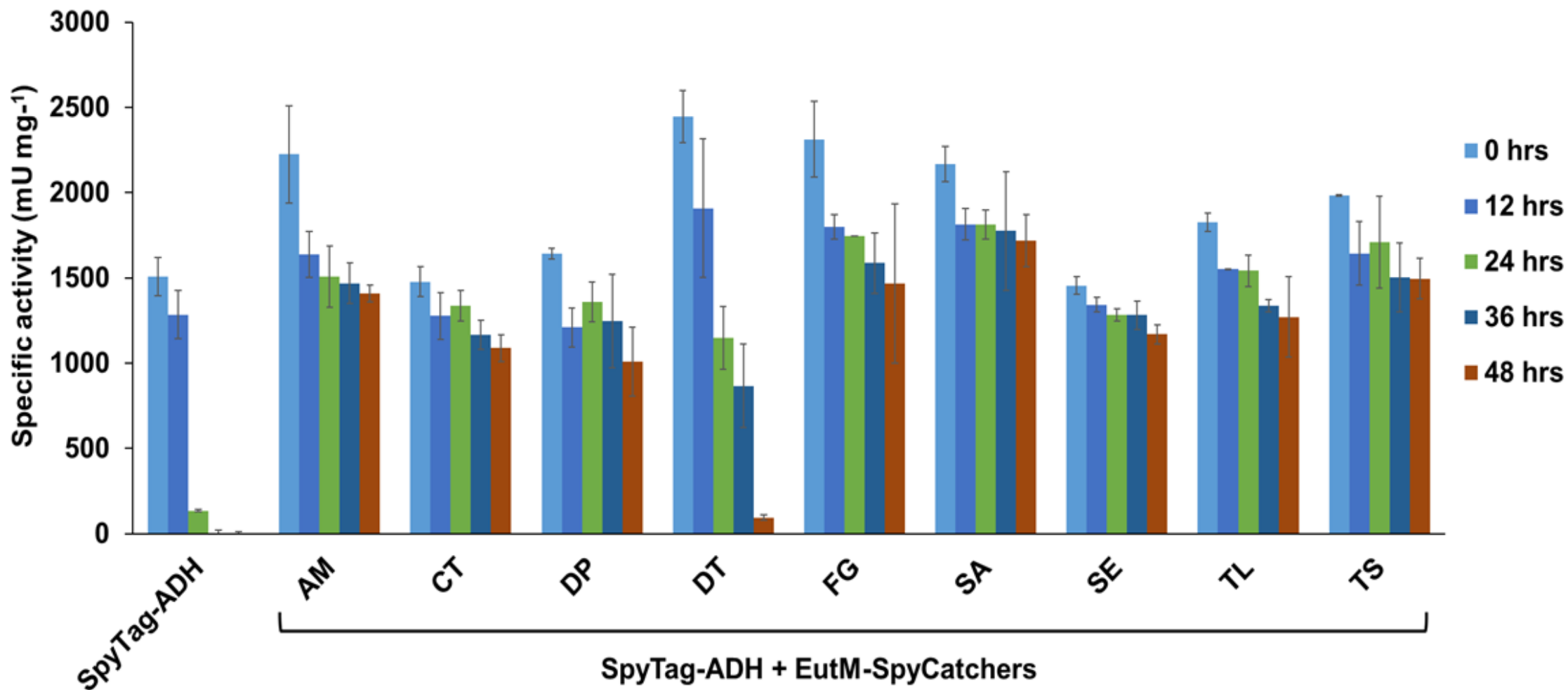
EutM TS

# Enzyme immobilization on EutM-SpyCatcher scaffolds



Enzyme activities measured with 20 mM (S)-2-hexanol (20 mM), 1 mM NAD<sup>+</sup> in 50 mM Tris-HCl (pH 8.0) of ADH (0.2 mg mL<sup>-1</sup>) in the absence (control) and presence of EutM-SpyCatcher scaffolds (at a 1:9 molar ratio).

# Enzyme immobilization on EutM-SpyCatcher scaffolds



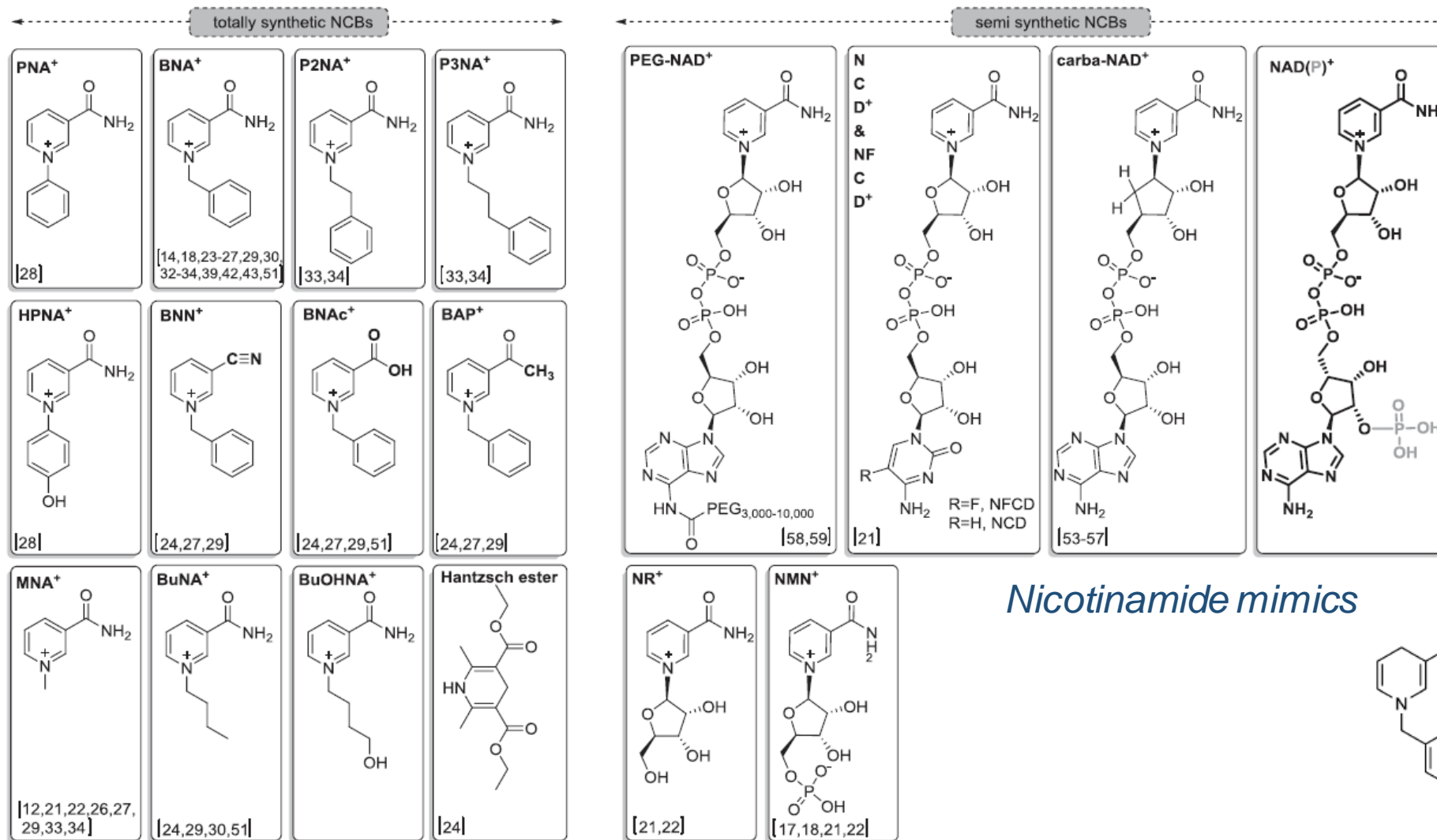
Enzyme (0.02 mg mL<sup>-1</sup>) mixed at a 1:9 molar ratio with EutM-SpyCatcher scaffolds in 50 mM Tris-HCl (pH 8.0) and incubated at 30°C for 0-48 hrs.

# ***Co-Factor stability & recycling***

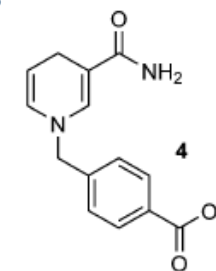
- Co-factor stability and recycling major limitation for the production of low- and medium-cost chemicals using biocatalysis due to the high cost of co-factors, especially nicotinamide adenine dinucleotide (NAD(P)) used for commonly used redox reactions in biocatalysis.
- Solutions to date – *In situ* co-factor generation systems and redox neutral and/or co-factor recycling enzyme cascades
- Co-factor stability problem remains ....

**Development of (acid-) stable, synthetic biomimetic co-factors to reduce process costs**

## Co-Factor stability & recycling



## Nicotinamide mimics

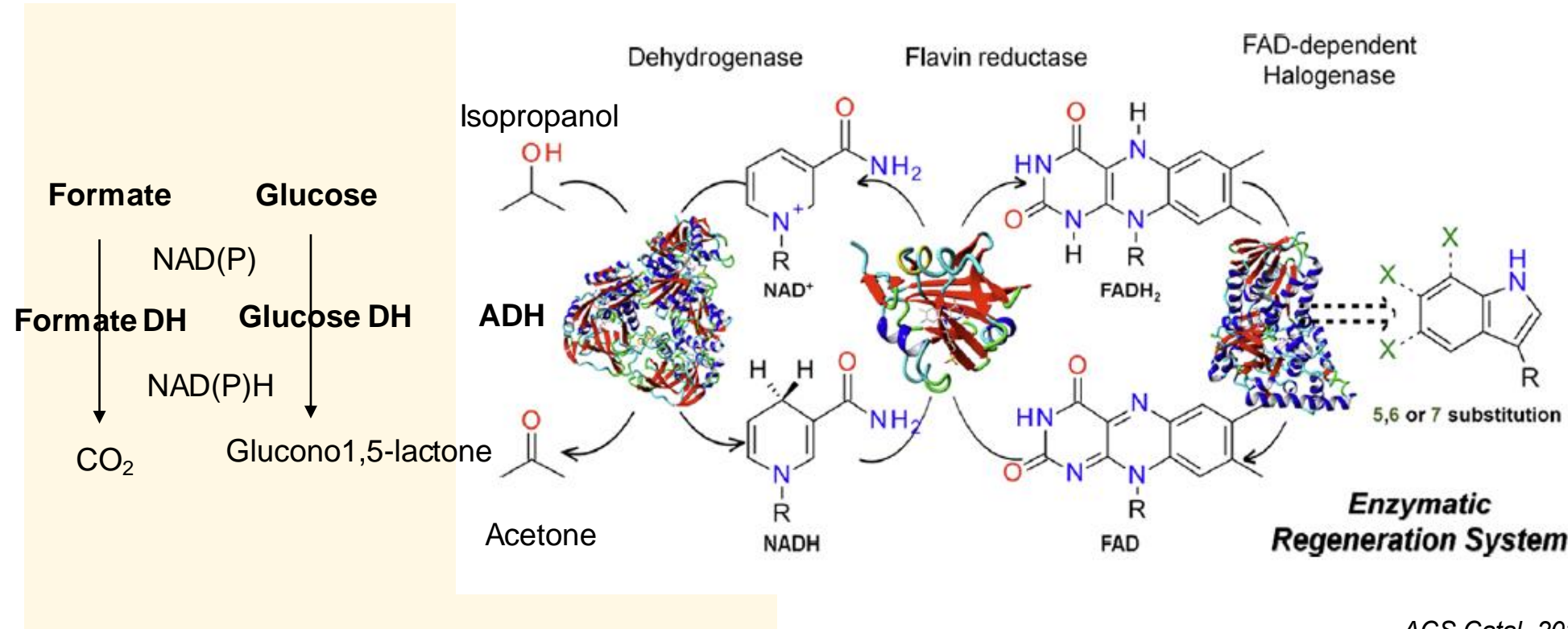


ChemBioChem 2019, 20, 838 – 845

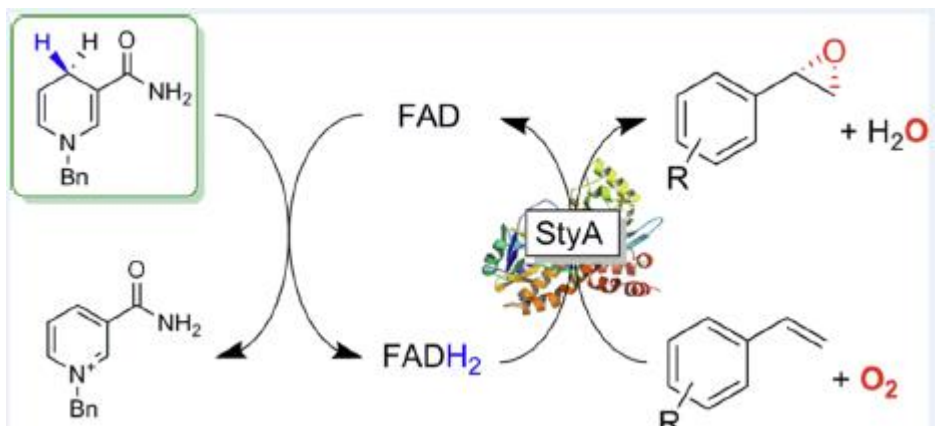


# Co-Factor stability & recycling

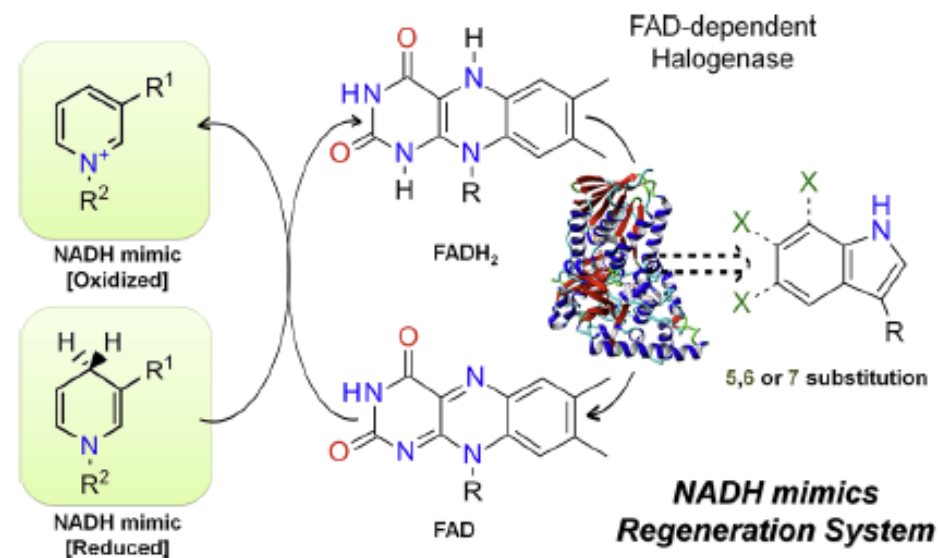
## FAD-dependent enzyme



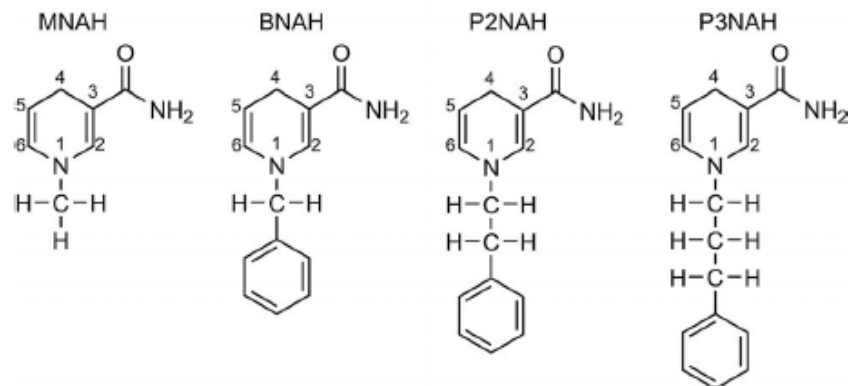
# Co-Factor stability & recycling



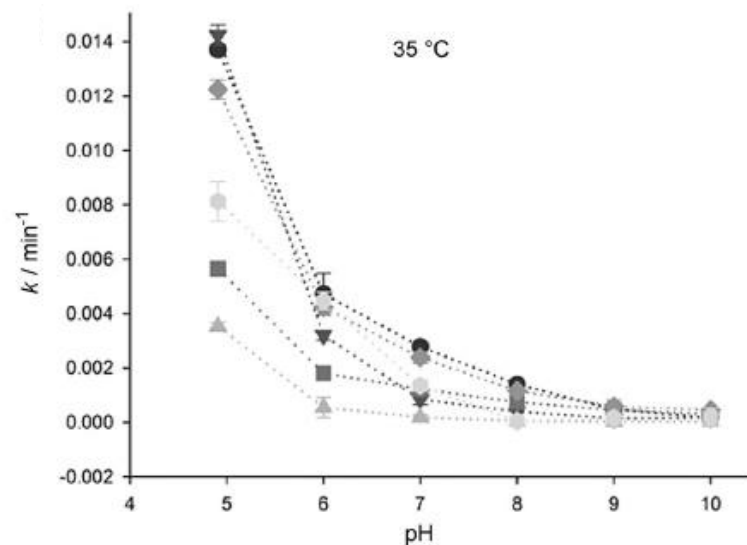
ACS Catal. 2015, 5, 2961–2965



ACS Catal. 2019, 9, 1389–1395



# Co-Factor stability & recycling



Mimics are not more stable,  
but cheaper to synthesize

Increasing number of examples show that mimics  
can yield comparable kinetics compared to natural  
NAD-cofactors

Limitations so far, lack of enzymatic in situ regeneration system  
for mimics & general applicability for oxidoreductases

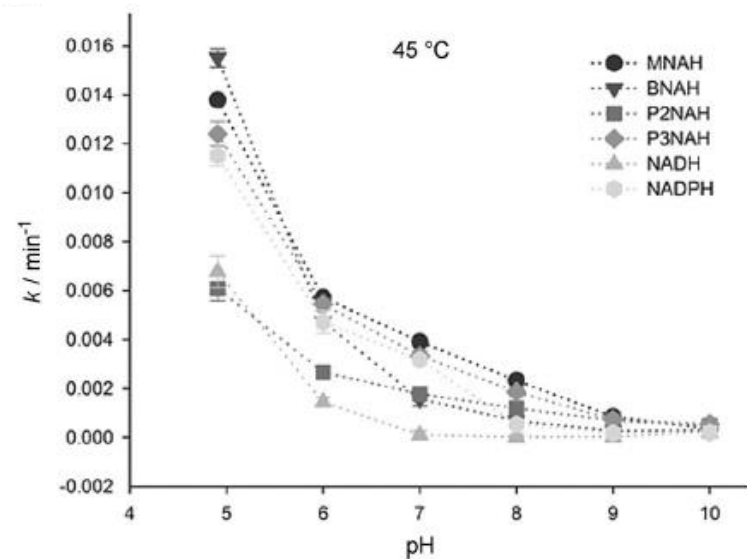
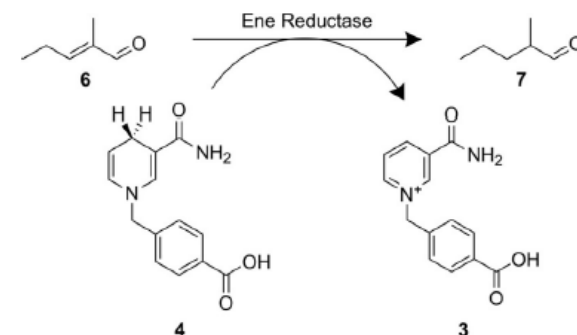


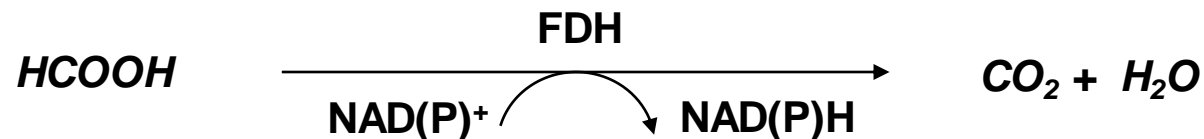
Table 2. The substrates and products used in ER-catalyzed reduction with compound 4 or NADH as the coenzyme, and the kinetic data for each substrate.

Substrate	Product	$K_m$ [mM]	Compound 4			NADH		
			$V_{max}$ [mU]	$k_{cat}/K_m$ [min <sup>-1</sup> ]	$K_m$ [mM]	$V_{max}$ [mU]	$k_{cat}/K_m$ [min <sup>-1</sup> ]	
		$0.07 \pm 0.016$	$15 \pm 0.04$	111.36	$0.03 \pm 0.0094$	$0.8 \pm 0.05$	11.62	
		$0.22 \pm 0.067$	$4 \pm 0.6$	7.97	$0.36 \pm 0.0503$	$0.9 \pm 0.04$	1.22	
		$0.26 \pm 0.046$	$3 \pm 0.07$	5.32	$0.48 \pm 0.299$	$0.6 \pm 0.04$	0.61	

$K_m$ : Michaelis–Menten constant,  $k_{cat}$ : turnover number [min<sup>-1</sup>] not shown.

# Energy & substrate input

*Electrons scavenged from glucose, formate, alcohols can drive biocatalytic reduction reactions*

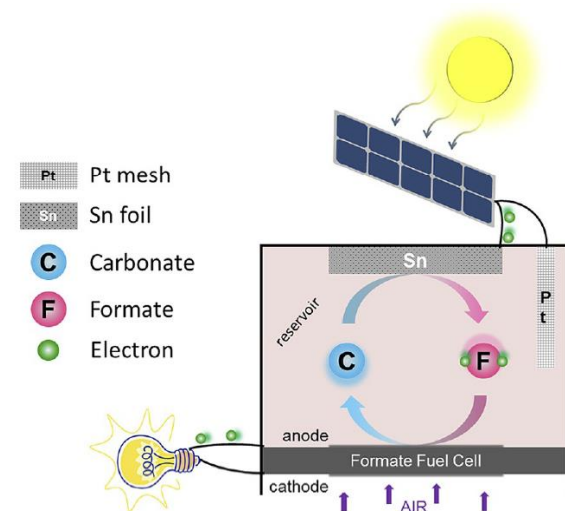


## Example – Formate as electron donor

### Formate: an Energy Storage and Transport Bridge between Carbon Dioxide and a Formate Fuel Cell in a Single Device

Tracy Vo, Krutarth Purohit, Christopher Nguyen, Brenna Biggs, Salvador Mayoral, and John L. Haan<sup>\*,[a]</sup>

ChemSusChem 2015, 8, 3853 – 3858



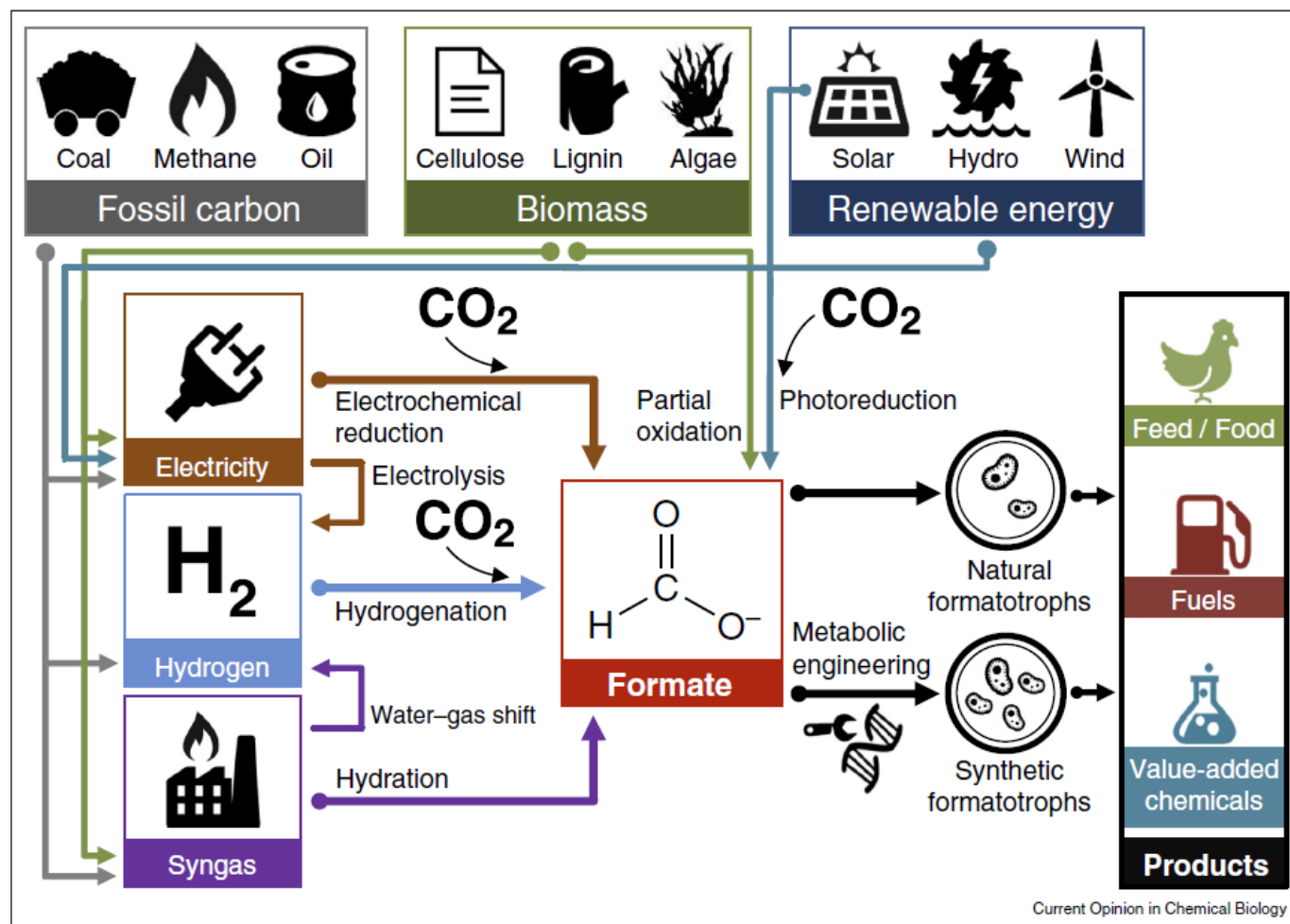
**Figure 6.** A 2D working principle of the device. Carbonate is converted to formate at a Sn electrode powered by a solar panel. The formate then “carries” the electrons to the DFFC where the formate is oxidized to release the electrons that perform external work. The oxidized formate diffuses back to the reservoir bulk to complete the cycle. Formate can be produced anytime energy is applied to the device, and power can be extracted anytime a load is applied.

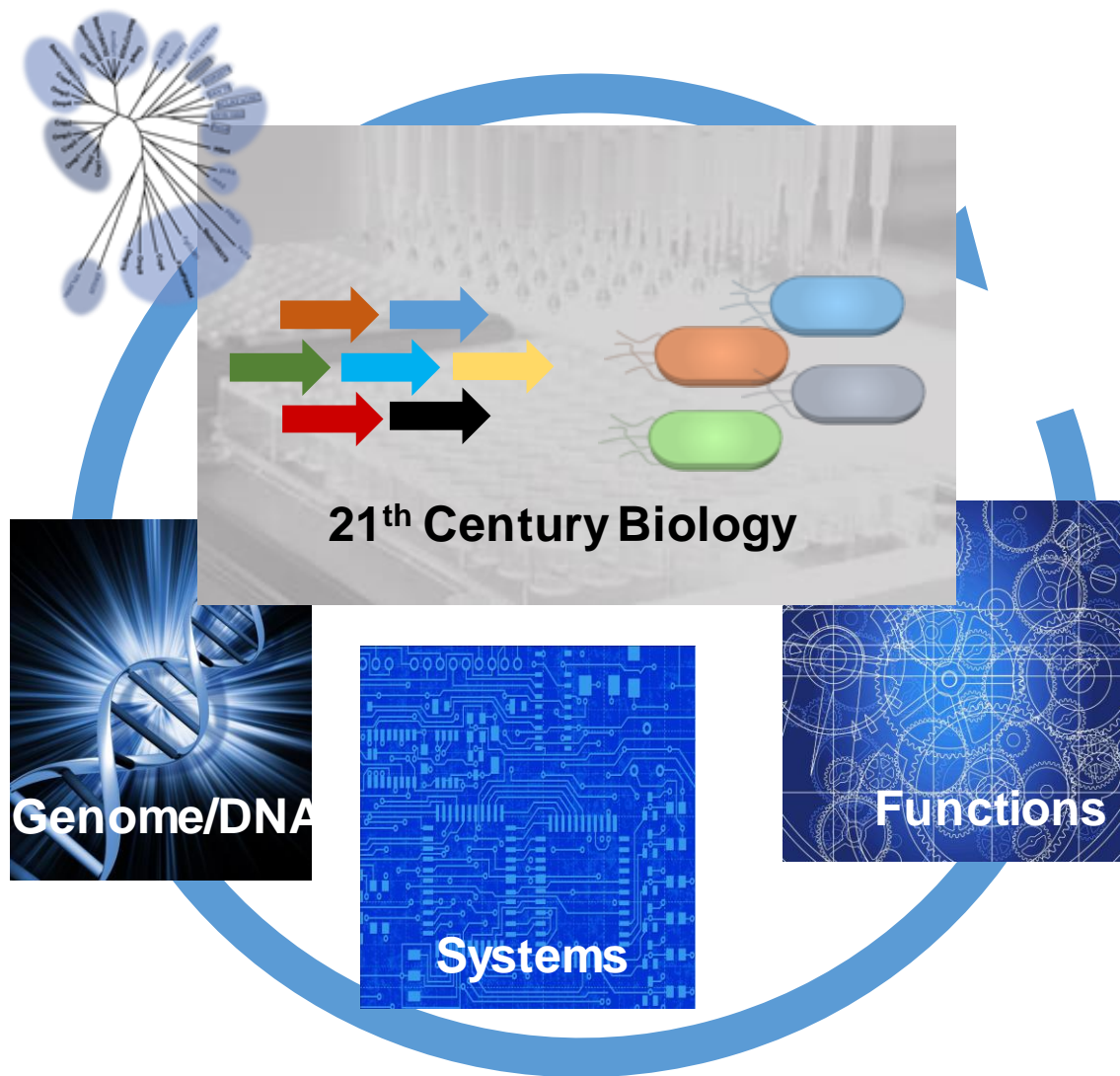
# Energy & substrate input

## The formate bio-economy

Oren Yishai, Steffen N Lindner, Jorge Gonzalez de la Cruz,  
Hezi Tenenboim and Arren Bar-Even

*Current Opinion in Chemical Biology* 2016, 35:1–9





## Biomanufacturing

